

Research

**Bacteriological profile and antibiotic sensitivity pattern of uropathogens in hemodialysis patients attending tertiary care hospital, Kathmandu, Nepal**

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**Abstract:** Hemodialysis (HD) patients are more susceptible to urinary tract infection (UTI) and UTIs are an important cause of morbidity and mortality in these patients. This study was conducted to find out the prevalence of UTI and multidrug-resistant (MDR) patterns of isolates among HD patients. A cross-sectional study was carried out from February 2018 to August 2018 at the National Kidney Center (NKC), Banasthali, Kathmandu. A total of 200 (108 male and 98 female) mid-stream urine samples were collected from patients with renal failure undergoing hemodialysis. Of them, 26% of samples showed significant bacteriuria. 22.2% and 30.4% of samples showed significant bacteriuria among males and females, respectively ( $p=0.19$ ). Among the age-group, bacteriuria was ranked top at the age group >70 years (35.3.8%) followed by 51-70 years (34.1%) ( $p=0.046$ ). Nine different bacteria were isolated. Among them, the most predominant organism was *Esch. coli* (32.7%) followed by *Staph. aureus* (26.9%), *Staph. saprophyticus* (13.5%) and others. For the Gram-negative isolates, Imipenem (96.8%) was found to be the most effective drug followed by Amikacin (83.9%) and Cefepime (64.5%). For Gram-positive isolates, Cefepime (76.1%) was found to be the most effective drug. The organisms showed 71.4% resistance to Cotrimoxazole and 57.1% resistance to Amoxicillin, Azithromycin, and Nitrofurantoin. The overall prevalence of MDR was found to be 57.7% in HD patients. Amikacin and Imipenem were found to be the drug of choice to treat UTI in HD patients. This study will be beneficial for making treatment policy and reducing the risk of UTI in HD patients.

**Keywords:** Hemodialysis, UTI, bacteriuria, multidrug-resistant, Nepal

## **Introduction**

Hemodialysis (HD) patients are more susceptible to bacterial urinary tract infection (UTI) and UTIs are an important cause of morbidity and mortality in these patients [1]. HD patients are at high risk for infections, which is attributed to impaired immune defenses, a high severity of illness, and the need for routine puncture of a vascular access site to remove blood for hemodialysis [2]. Moreover, urine voiding is a natural process of bacterial clearance in the urinary tract [3]. Usually, HD patients are subjected to excrete urine mechanically and lack of natural voiding to excrete urine is often overlooked as a source of infection in HD patients [4]. UTI is the second most common cause of death in patients dependent on HD [5]. Therefore, routine monitoring and diagnosis of urinary tract infection (UTI) on time is very crucial that help in infection control and therapeutic management [4, 6].

Mounting evidence has already demonstrated that HD patients are more vulnerable to UTI. The prevalence of pyuria in dialysis patients from different studies ranges between 28% and 72% and the prevalence of documented UTI in those studies varied between 11% and 70% [7]. A very high incidence of UTI was reported by Jadav et al (1977) [8] both in acute renal failure (73.0%) and chronic renal failure patients (57.5%) in Mumbai (India). Likewise, in a study conducted in Nepal in chronic HD patients, Gram-negative organisms were isolated at a rate of 84.6% [9]. In another study, 21.1% were found to have a final diagnosis of UTI on urine discharge [10]. Amikacin, Imipenem, Ceftazidime, and Gentamicin are antibiotics of choice for the treatment of UTI in HD patients.

This study was aimed to determine the bacterial profile and antimicrobial susceptibility pattern of organisms causing urinary tract infections in hemodialysis patients and detect Multi-Drug Resistant (MDR) producing strains of isolates.

## **Materials and methods**

### **Study area and sample size**

A cross-sectional study was conducted in the National Kidney Center's Laboratory, Kathmandu, Nepal from Feb 2018 to Aug 2018. The sample size was determined by Fisher's Formula. 200 urine samples were taken from patients with renal failure undergoing hemodialysis. 5-10 ml clean-catch midstream urine was collected in a leak-proof, wide-mouthed, properly capped, sterile container in the hospital on the day of dialysis. The urine sample was transported to the laboratory

as soon as possible for further process. The urine was cultured onto the MacConkey agar and blood agar medium by the semi-quantitative culture techniques using a standard loop. Isolates were identified on the basis of standard morphological appearance of the colonies, staining reactions, biochemical properties and serotyping if required in specific cases [11, 12]. *Staph. saprophyticus* was identified by Novobiocin test.

### Bacterial Antibiotic Sensitivity Test

Antimicrobial susceptibility testing of the isolates towards various antimicrobial disks was done by the modified Kirby-Bauer disk diffusion method as recommended by CLSI 2014 using Mueller Hinton Agar. The antibiotics used were amikacin, azithromycin, cefepime, ceftriaxone, amoxicillin, ciprofloxacin, levofloxacin, cotrimoxazole, nitrofurantoin, chloramphenicol, and linezolid. Based on the susceptibility pattern of isolates, bacteria resistant to at least more than 2 classes of antibiotics were considered as Multi-Drug Resistant [13].

### Ethical approval

Ethical clearance was obtained from the Institutional Review Committee of Shi-Gan Health Foundation and National Institute of Tropical Medicine, Kathmandu prior to the research (IRC Reference No: 04/2071/08/26).

## Results

### Growth pattern in two genders:

Out of the total 200 urine samples, 26% (52/200) of samples showed significant bacteriuria. A marginally higher positive rate was seen in females (30.4%; 28/98) compared to their male counterparts (22.2%; 24/108) ( $p=0.19$ ) (Table 1).

Table 1: Bacterial growth positivity in hemodialysis patients of two genders.

Sex	Total count (n)	Positive count (n)	%	P-value
Male	108	24	22.2%	0.19
Female	98	28	30.4%	
<b>Total</b>	<b>200</b>	<b>52</b>	<b>26.0</b>	

Among HD patients of different age groups, highest growth positive rate was observed in the age-group of more than 70 years followed by 51 – 70 years (35.3%; 6/17) and 31 – 50 years (34.1%; 30/88) and the least growth positive rate was seen in the age group of 11-30 years (15.4%; 4/26).

This increasing trend with age was statistically significant ( $p=0.046$ ) (Table 2). In males, the highest rate of growth positive was seen in the age-group of  $>70$  (33.3%) and lowest in the age-group 11-30 (6.7%) while in females, highest significant bacteriuria was seen in the age-group 51-70 (47.1%) and the least in the age-group 31-50 (13.9%) (Table 2).

Table 2: Bacterial growth positivity in HD patients of different sex and age-groups.

Age group	Male		Female		Total	
	Total (n)	Growth positive	Total (n)	Growth positive	Total (n)	Growth positive
11-30	15	1 (6.7%)	11	3 (27.3%)	26	4 (15.4%)
31-50	33	7 (21.2%)	36	5 (13.9%)	69	12 (17.4%)
51-70	54	14 (25.9%)	34	16(47.1%)	88	30 (34.1%)
$>70$	6	2 (33.3%)	11	4 (36.4%)	17	6 (35.3%)
<b>Total</b>	<b>108</b>	<b>24 (22.2%)</b>	<b>92</b>	<b>28 (30.3%)</b>	<b>200</b>	<b>52 (26.00%)</b>

### Bacterial profile and multi-drug resistant strain

A total of 52 bacterial strains were isolated from 200 HD patients. *Escherichia. coli* was the most frequently isolated bacterial spp. Of 52 bacterial isolates, 30 (57.7%) were MDR. Among MDR strains, *Staph. aureus* (78.6%) isolates were most predominant MDR (Table 3).

Table 3. Multidrug-resistant strain isolated from HD patients.

Organisms	Total isolates (n)	MDR strains (%)
<i>Escherichia coli</i>	17	7 (41.2%)
<i>Staphylococcus aureus</i>	14	11 (78.6%)
<i>Staphylococcus saprophyticus</i>	7	5 (71.4%)
<i>Klebsiella oxytoca</i>	5	2 (40.0%)
<i>Citrobacter freundii</i>	4	2 (50.0%)
<i>Others</i>	5	3 (60.0%)
<b>Total</b>	<b>52</b>	<b>30 (57.7%)</b>

### Antibiotic sensitivity pattern of Gram-positive isolates

For Gram-positive isolates, Cefepime (76.1%) was found to be a more effective drug. The Gram-positive isolates showed 71.4% resistance to Cotrimoxazole and 57.1% resistance to Amoxicillin, Azithromycin, and Nitrofurantoin (Table 4).

### Antibiotic sensitivity pattern of Gram-negative isolates

Among the Gram-negative isolates, Imipenem (96.8%) was found to be the most effective drug followed by Amikacin (83.9%) and Cefepime (64.5%). The Gram-negative isolates showed 54.8% resistivity to Cephalexin and Ciprofloxacin (Table 5).

Table 4: Antibiotic sensitivity pattern of Gram-positive isolates.

Antibiotics used	Susceptibility pattern		
	Resistant	Intermediate	Sensitive
	n (%)	n (%)	n (%)
Amikacin	5 (23.8%)	1 (4.8%)	15 (71.4%)
Azithromycin	12 (57.1%)	6 (28.6%)	3 (14.3%)
Cefepime	4 (19.1%)	1 (4.8%)	16 (76.1%)
Ceftriaxone	11 (52.4%)	2 (9.5%)	8 (38.1%)
Amoxicillin	12 (57.1%)	6 (28.6%)	3 (14.3%)
Ciprofloxacin	8 (38.1%)	9 (42.9%)	4 (19.1%)
Levofloxacin	9 (42.8%)	6 (28.6%)	6 (28.6%)
Cotrimoxazole	15 (71.4%)	3 (14.3%)	3 (14.3%)
Nitrofurantoin	12 (57.1%)	6 (28.6%)	3 (14.3%)
Chloramphenicol	2 (9.5%)	5 (23.8%)	14 (66.7%)
Linezolid	6 (28.6%)	2 (9.5%)	13 (61.9%)

n = 21

Table 5: Antibiotic sensitivity pattern of Gram-negative isolates

Antibiotics used	Susceptibility pattern		
	Resistant	Intermediate	Sensitive
	n (%)	n (%)	n (%)
Amikacin	0 (0.0%)	5 (16.1%)	26 (83.9%)
Azithromycin	16 (51.6%)	6 (19.4%)	9 (29.0%)
Cefepime	7 (22.6%)	4 (12.9%)	20 (64.5%)
Ceftriaxone	10 (32.3%)	6 (19.4%)	15 (48.3%)
Cephalexin	17 (54.8%)	1 (3.2%)	13 (41.9%)
Imipenem	0 (0.0%)	1 (3.2%)	30 (96.8%)
Ciprofloxacin	17 (54.8%)	6 (19.4%)	8 (25.8%)
Levofloxacin	4 (12.9%)	9 (29.0%)	18 (58.1%)
Norfloxacin	14 (45.2%)	7 (22.6%)	10 (32.1%)
Ofloxacin	13 (41.9%)	7 (22.6%)	11 (35.5%)
Cotrimoxazole	13 (41.9%)	5 (16.2%)	13 (41.9%)
Nitrofurantoin	14 (45.2%)	7 (22.6%)	10 (32.2%)

n = 31

## Discussion

In this study, one-fourth of the samples showed significant bacteriuria indicating the cases of UTI. This finding is lower than the finding of Singh et al (2016) [14] which was 34.0% and higher than the finding of Pradhan and Pradhan (2017) [15] (13.8%). However, similar finding has been reported by Dhakal et al (1999) [16] (25.2%). A marginally higher culture-positive rate was observed in females (30.4%) than in their male counterparts (22.2%). The higher rate of UTI in females was also reported by Pardeshi (2018), Daoud and Afif (2011), Khatiwada et al (2018) and Jha and Bapat (2005) [17, 18, 19, 20]. This might be mainly due to the anatomical and behavioral differences among females.

UTI in HD patients included in this study showed an increasing trend with age (15.4% in the age group of 11-30 years to 35.3% in the age group of >70 years) and this increment was significantly higher. Haider et al (2017) also reported a higher prevalence of UTI in the elderly groups [21]. This might be mainly due to the increasing prevalence of risk factors such as diabetes, hypertension and cardiovascular disease among older groups [22]. In the United States, chronic kidney disease was reported to be about 10% in patients of age >65 years, in contrast to 1.5% of the younger employed population [23].

Of the different kinds of pathogens isolated in this study *Esch. coli* was the commonest (17/52; 32.7%) followed by *Staph. aureus* (14/52; 26.9%) and *Staph. saprophyticus* (7/52; 13.5%) and others. The overall multi-drug resistance in this study was found to be 57.7% (30/52). *Staph. aureus* showed the highest degree of MDR (78.6%) followed by *Staph. saprophyticus* (71.4%). Different rates of prevalence of MDR ranging as low as 3% to as high as 59% in hemodialysis have been reported by other investigators [9, 24, 25, 26, 29]. This might be associated with different risk factors such as prior hospitalization, temporary dialysis access, residence in nursing homes, and antimicrobial exposure [24, 25, 27, 28].

Gram-negative bacilli (59.6%) were the predominant organisms isolated from urine samples than the Gram-positive cocci (40.4%). This finding is in agreement with those of Moges et al (2002) [30]; Kothari and Sagar (2008) [31]; Puri et al (2006) [32] and Karki et al (2004) [33]. Among the Gram-negative bacteria, *Esch. coli* was found as the most predominant organism. This result is in agreement with those of Beyene and Tsegaye (2011) [34]; Daoud and Afif (2011) [18]; Elkehili et al (2010) [35]; Aypak et al (2009) [36] and Rai et al (2008) [37]. The major factor responsible for

the high prevalence of uropathogenic *Esch. coli* (UPEC) is the type 1-pilli which enhances binding and invasion to the superficial epithelial cells [38].

*Staph. aureus* was found as the second most predominant organism in this study. This finding is different than the findings of Chaudhary et al (2016) [9]. The presence of this organism in urine often indicates pyelonephritis acquired via a hematogenous route or descending route. This finding, however, is in agreement with the findings of Nicholas et al (2018) [39] where *Staph. aureus* is the predominant organism in HD patients to cause septicemia.

In this study, Imipenem (96.8%) was found to be the most effective drug against Gram-negative bacterial isolates followed by Amikacin (83.9%) which is similar to the study done by Khatiwada et al [19]. All *Esch. coli* isolates were susceptible to Imipenem followed by Amikacin (94.1%). Quinolone/ Fluoroquinolone (Ciprofloxacin, Norfloxacin) were found least active. A study done in Senegal has shown that the increasing resistance to these drugs is hypothesized for the generalized use of fluoroquinolones in animal feed and subsequent transmission of resistant strains from animals to humans [40]. *Acinetobacter* spp. showed 100% sensitivity to aminoglycosides, macrolide and  $\beta$ -lactam groups of antibiotics used. *Ps. aeruginosa* showed 100% resistivity to Nitrofurantoin and Cephalexin. Several factors are responsible for the rise of resistance rate of bacterial uropathogens including misuse of antibiotics, frequent oral use of wide-spectrum antimicrobials that may change intestinal flora (which is usually common cause of UTI) and inappropriate dosage and duration of treatments [41]. Though aminoglycosides are effective against Gram-negative bacteria in this study, the use of aminoglycosides in HD patients should be assessed against their ototoxicity and nephrotoxicity [42].

## Conclusion

The study findings showed that *Esch. coli* were the most predominant pathogens among the HD patients followed by *Staph. aureus* with an increasing trend with age. This study also showed a marginally higher prevalence of UTI in female HD patients than in their male counterparts. The study showed that Imipenem and Amikacin were the most effective drug for Gram-positive bacteria and Imipenem and Amikacin for Gram-negative bacteria with the overall prevalence of MDR of 57.7%. Our data will be beneficial for making treatment policy and reducing the risk of UTI in HD patients.



## References

1. Fasolo LR, Rocha LM, Campbell S and Peixoto AJ (2006). Diagnostic relevance of pyuria in dialysis patients. *Kidney Int.* **70**: 2035–2038.
2. Horl WH (1999). Neutrophil function and infections in uremia. *Am J Kidney Dis.* **33**(2): 45-47.
3. Abraham SN and Miao Y (2015). The nature of immune responses to urinary tract infections. *Nat Rev Immunol.* **15**(10): 655-663.
4. Bennett WM and Craven R (1976). Urinary tract infections in patients with severe renal disease. Treatment with ampicillin and trimethoprim-sulfamethoxazole. *JAMA.* **236**(8): 946–948.
5. Sarnak MJ and Jaber BL (2000). Mortality caused by sepsis in patients with end-stage renal disease compared with the general population. *Kidney Int.* **5**: 1754-1764.
6. D'Agata EM, Mount D, Thayer V and Schaffner W (2000). Hospital-acquired infections among chronic hemodialysis patients. *JAMA.* **35**(6): 1083-1088.
7. Fasolo LR, Rocha LM, Campbell S and Peixoto AJ (2006). Diagnostic relevance of pyuria in dialysis patients. *Kidney Int.* **70**(11): 2035 – 2038.
8. Jadav SK, and Sant SM (1977). Bacteriology of urinary tract infection in patients of renal failure undergoing dialysis. *Postgrad Med J.* **23**(1): 10-18.
9. Chaudhary R, Chaturvedi SB, Shah PK, Pant AD and Pokharel N (2016). Bacteriology of Urinary Tract Infection of Chronic Renal Failure Patients Undergoing for Hemodialysis. *J Microbiol Exp.* 3. 10.15406/jmen.2016.03.00089
10. Oikonomou KG and Alhaddad A (2016). The Diagnostic Value of Urinalysis in Hemodialysis Patients with Fever, Sepsis or Suspected Urinary Tract Infection. *J Clin Diagn Res.* **10**(10): 11–13.
11. Cheesebrough M (2012). District Laboratory Practice in Tropical Countries, New Delhi. Cambridge University Press **2**: 105-115.
12. Forbes BA, Sahm DF and Weissfeld AS (2007). Bailey and Scott's Diagnostic Microbiology, St. Louis, MO: Elsevier Mosby **12**: 843-845.
13. Clinical Laboratory Standards Institute (CLSI). CLSI document M100-S23. Performance standards for antimicrobial susceptibility testing: Twenty third informational supplemented. Wayne: CLSI; 2013.



14. Singh N, Gandhi S, McArthur E, Moist L, Jain AK and Liu AR (2015). Kidney function and the use of nitrofurantoin to treat urinary tract infections in older women. *Can Med Assoc.* **187**: 648-656.
15. Pradhan B and Pradhan SB (2017). Prevalence of Urinary Tract Infection and Antibiotic Susceptibility Pattern to Urinary Pathogens in Kathmandu Medical College and Teaching Hospital, Duwakot. Birat *J Health Sci.* **2**(1): 134-137.
16. Dhakal BK, Pokharel BM and Basnyat SR (1999). A prospective study of urinary tract infection based on culture and direct microscopy of urine along with the antibiotic sensitivity test of urinary pathogen. A dissertation submitted to the Central Department of Microbiology, TU, Kathmandu, Nepal.
17. Pardeshi P (2018). Prevalence of urinary tract infections and current scenario of antibiotic susceptibility pattern of bacteria causing UTI. *Indian J Microbiol Res.* **5**(3): 334-338.
18. Daoud Z and Afif C (2011). *E. coli* isolated from urinary tract infection of Lebanese patients between 2000 and 2009. Epidemiology and profile of resistance. Himdhai Publishing Corporation, Doi: 10.1155/2011/218431.
19. Khatiwada S, Khanal R, Karn SL, Raut S and Poudel A (2018). Antimicrobial susceptibility profile of urinary tract infection: A single-center hospital-based study from Nepal. *J Univ Coll Med Sci.* **6**(2): 37-40.
20. Jha N and Bapat SK (2005). A study of sensitivity and resistance of pathogenic microorganisms causing UTI to Kathmandu valley. *Kathmandu Univ Med J.* **3**:123-129.
21. Haider JS, Hasan A, Bin-Tahir K (2016) Frequency of Urinary Tract Bacterial Infection and their Susceptibility Patterns among Hemodialysis Patients in Zliten Hospital. *J Microbiol Exp.* **3**(3): 00093. DOI: 10.15406/jmen.2016.03.00093
22. Williams M (2013). Diabetic kidney disease in elderly individuals. *Med Clin N Am.* **97**(1): 75–89.
23. US Renal Data System. USRDS 2013. Atlas of Chronic Kidney Disease and End-Stage Renal Disease in the United States. National Institutes of Health, National Institute of Diabetes and Digestive and Kidney Diseases; Bethesda, MD, USA: 2013.
24. Zacharioudakis IM, Zervou FN, Ziakas PD, Rice LB, Mylonakis E (2015). Vancomycin-resistant enterococci colonization among dialysis patients: A meta-analysis of prevalence, risk factors, and significance. *Am J Kidney Dis.* **65**, 88–97.

25. Zacharioudakis IM, Zervou FN, Ziakas PD, Rice LB, Mylonakis E (2014). Meta-analysis of methicillin-resistant *Staphylococcus aureus* colonization and risk of infection in dialysis patients. *J Am Soc Nephrol.* **25**: 2131–2141.
26. Song JU, Park HK, Kang HK, Lee J (2017). Proposed risk factors for infection with multidrug-resistant pathogens in hemodialysis patients hospitalized with pneumonia. *BMC Infect Dis.* **17**: 681–690.
27. Karanika S, Zervou FN, Zacharioudakis IM, Paudel S, Mylonakis E (2015). Risk factors for methicillin-resistant *Staphylococcus aureus* colonization in dialysis patients: A meta-analysis. *J Hosp Infect.* **91**: 257–263.
28. Pop-Vicas A, Strom T, D'Agata EMC (2008). Multidrug-resistant gram-negative bacilli among chronic hemodialysis patients. *J Clin Microbiol.* **3**: 752–758.
29. Grabsch EA, Burrell LJ, Padiglione A, O'Keeffe JM, Ballard S, Grayson ML (2006). Risk of environmental and healthcare worker contamination with vancomycin-resistant enterococci during outpatient procedures and hemodialysis. *Infect Control Hosp Epidemiol.* **27**(3): 287-293.
30. Moges AF, Genetu A and Mengistu G (2002). Antibiotic sensitivity of common bacterial pathogens in urinary tract infections at Gondar Hospital Ethiopia. *East Afr Med J.* **76**(3):140-142.
31. Kothari A and Sagar V (2008). Antibiotic in pathogens causing community-acquired urinary tract infection in India: A multicenter study. *J Infect Dev Ctries.* **2**(5): 354-358.
32. Puri N, Jha B, Lekhak B and Adhikari RC (2006). Study on the incidence of urinary tract infection in diabetic patients and the prevalence of multidrug-resistant strains among the bacteria isolates. A dissertation submitted to the Central Department of Microbiology, Tribhuvan University, Kathmandu, Nepal.
33. Karki A, Tiwari BR, and Pradhan SB (2004). Study of bacterial isolated from urinary tract infection and their sensitivity pattern. *J Nepal Med Assoc.* **43**: 200-203.
34. Beyene G and Tsegaye W (2011). Bacterial uropathogens in Urinary Tract Infection and antibiotic susceptibility pattern in Jimma University Specialized Hospital, Ethiopia. *J Health Sci.* **12**(2): 141-146.
35. Elkehili IM, Kekli AB, Zaak AS and Salem EL (2010). Urinary tract infection renal transplant recipients. *Arab J Nephrol Transplant.* **3**(2): 53-55.

36. Aypak C, Adalet AA and Duzgun N (2009). Empiric antibiotic therapy in acute uncomplicated urinary tract infection and fluoroquinolone resistance: a prospective observational study. *Ann Clin Microbiol Antimicrob.* **8**:27. doi: 10.1186/1476-0711-8-27.
37. Rai GK, Upreti HC, Rai SK, Shah KP and Shrestha RM (2008). Causative agent of urinary tract infection in children and their antibiotic sensitivity pattern: a hospital-based study. *Nepal Med Coll J.* **10**(2): 86-90.
38. Peerayeh S, Navidinia M, Fallah F and Jamali J (2018). Pathogenicity determinants and epidemiology of uropathogenic *E. coli* (UPEC) strains isolated from children with urinary tract infection (UTI) to define distinct pathotypes. *Biomed Res.* **29**. 10.4066/biomedicalresearch.29-17-1591.
39. Nicholas V, Neda F and Kathryn F (2018). Rate of bacteremia in the hemodialysis patient presenting to the emergency department with fever: a retrospective chart review. *Int Emerg J Med.* **29**(11): 1-6.
40. Sire JM, Nabeth P, Claude JDP, Bahsoun I, Siby T, Macondo, EA, Diallo AG, Gayomand S, Seek A, Breurec S and Grin B (2007). Antimicrobial resistance to outpatient *E. coli* urinary isolates in Dakar, Senegal. *J Infec Dev Ctries.* **1**(3): 263-268.
41. Shara MA (2011). A five-year etiology and antimicrobial susceptibility pattern of urinary pathogens in children at Princess Rahmah Hospital, Jordan. *Saudi J Kidney Dis Transpl.* **22**(6): 1249-1252.
42. Manhal FS, Mohammed AA and Ali KH (2012). Urinary tract infection in Hemodialysis patients with renal failure. *J Fac Med Baghdad.* **54**(1): 38-41.

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## Author's contribution

**Miya, A.K.** designed and performed experiments. **Ansari, M.** designed the study, analyzed the data and wrote the manuscript. **Rai, G.** revised and approved the work plan, supervised the overall work and helped in data analysis. **Pant, A.D.** helped in experimental design and

supervised the lab work. **Rai, K.R** wrote and revised the manuscript and helped in data analysis. **Rai, S.K.** revised the work plan, experimental design, and manuscript. All authors read and approved the final manuscript.

### **Dedication**

Not mentioned

### **Conflict of Interest**

There are no conflicts to declare.

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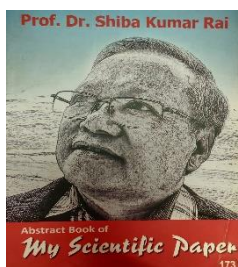
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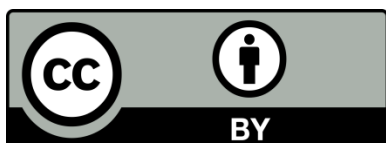
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